

Supplementary Information For:

MacP bypass variants of *Streptococcus pneumoniae* PBP2a suggest a conserved mechanism for the activation of bifunctional cell wall synthases.

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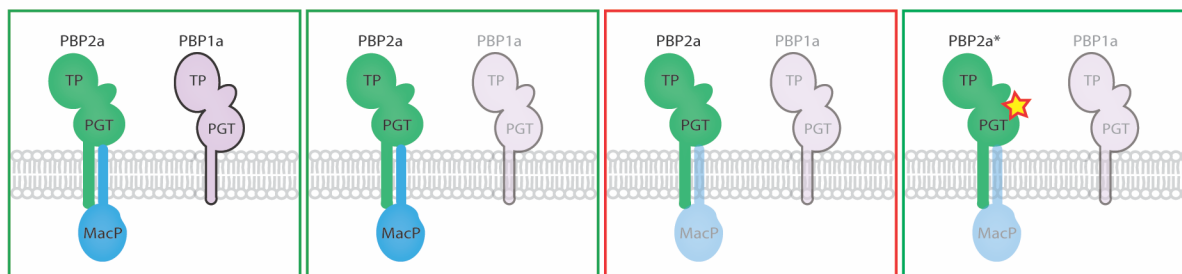
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A



B

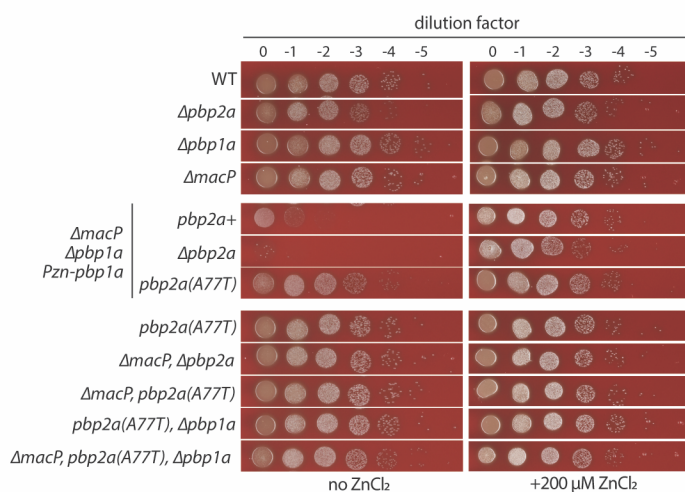
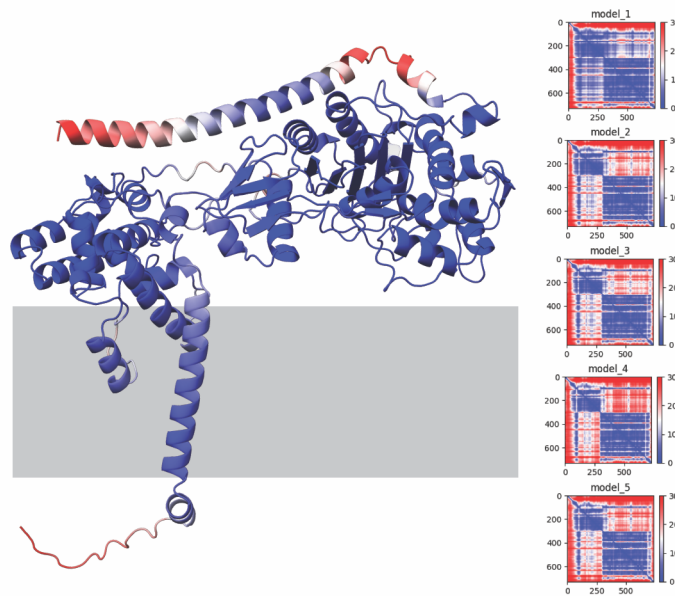


Figure S1. Schematic representation of the selection for PBP2a mutants that bypass the requirement for MacP. (A) Boxes depict viable (green) and non-viable (red) strains. PPB1a depletion and *macP* deletion are represented as translucent shapes. PBP2a mutants that bypass the requirement for MacP are shown with a yellow star. **(B)** Representative spot dilutions of the indicated strains on blood agar plates in the absence and presence of 200 μM ZnCl₂. The data in the figure are from one of three biological replicates.

A

PBP2a mutants tested for growth in $\Delta macP$, $\Delta pbp1a$, $Pzn-pbp1A$ background													
A75	V76	A77	K78	S79	T80	D84	K90	R92	D97	R98	E99	E100	K101
A75T	V76T	A77C	K78A	S79A	T80A	D84K	K90D	R92D	D97K	R98D	E99K	E100K	K101D
		A77D	K78D	S79V	T80V			R92E			E99R	E100R	
		A77E											
		A77F											
		A77G											
		A77H											
		A77I											
		A77K											
		A77L											
		A77M											
		A77N											
		A77P											
		A77Q											
		A77R											
		A77S											
		A77T											
		A77V											
		A77W											
		A77Y											

B



C

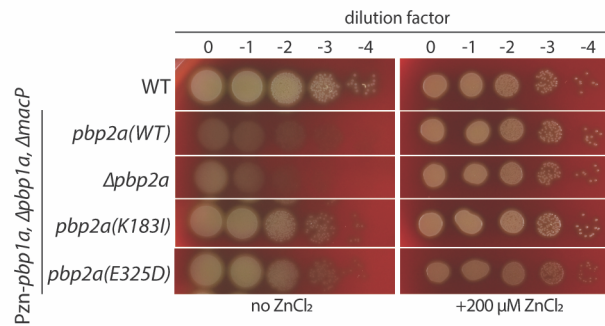


Figure S2. PBP2a variants tested for the ability to bypass MacP

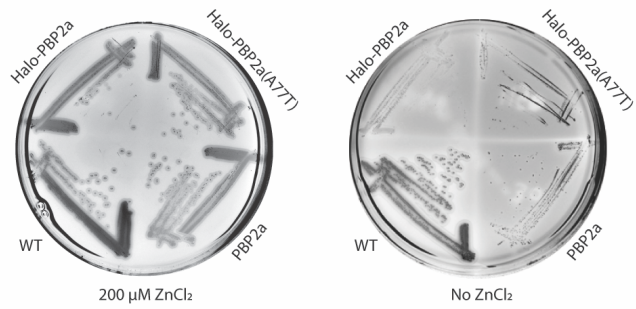
(A) Table of the PBP2a variants generated by site-directed mutagenesis and tested for the ability to support growth of a $\Delta macP$ mutant upon depletion of PBP1a. Each mutant was tested for growth in strain Sp1047 ($\Delta pbp1a$, $\Delta macP$, $Pzn-pbp1a$, $\Delta pbp2a$) by introducing the indicated *pbp2a* allele. Mutants that were unable to sustain growth when PBP1a was depleted are shown in black. Mutants that could bypass MacP are shown in

green. **(B)** Structural model of PBP2a with predicted local distance difference test (pLDDT) for all residues in the predicted structure. Low (red) to high (blue) confidence is indicated. The predicted alignment error (pAE) plots for the top 5 models generated. **(C)** Representative spot dilutions of the indicated strains on blood agar plates in the absence and presence of $ZnCl_2$. Mutants K183I and E325D also bypass MacP requirements. Data in panel C are from one of three biological replicates.

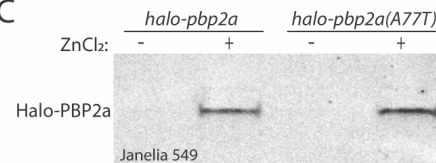
A

1st replicate	WT	Δ pbp2a	pbp2a(A77T)	Δ bp1a, Δ macP, pbp2a(A77T)
PBP2a signal	7568	1	4069	10336
n.s. signal	8579	9789	10864	10606
ratio	0.88	0	0.37	0.97
normalized with WT	1	0	0.42	1.1
2nd replicate	WT	Δ pbp2a	pbp2a(A77T)	Δ bp1a, Δ macP, pbp2a(A77T)
PBP2a signal	3633	0	2088	10736
n.s. signal	3500	4352	5058	8918
ratio	1.04	0	0.41	1.2
normalized with WT	1	0	0.4	1.16
3rd replicate	WT	Δ pbp2a	pbp2a(A77T)	Δ bp1a, Δ macP, pbp2a(A77T)
PBP2a signal	5964	0	4302	8408
n.s. signal	9258	9629	9337	10593
ratio	0.64	0	0.46	0.79
normalized with WT	1	0	0.72	1.23

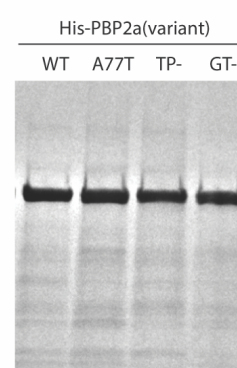
B



C



D



E

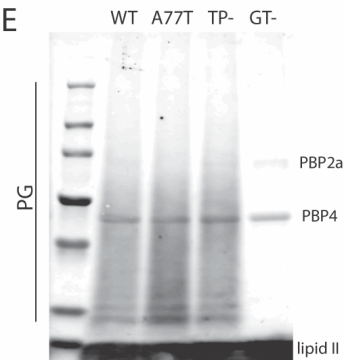


Figure S3. Analysis of PBP2A *in vivo* and *in vitro*.

(A) Table documenting the relative amounts of PBP2a in the indicated strains. Immunoblots from three biological replicates were analyzed, and the intensities (arbitrary units) of PBP2a and the non-specific (n.s.) bands were quantified using ImageJ. No significant enrichment was observed for Δ pbp1a, Δ macP, or pbp2a(A77T) strains. However, PBP2a(A77T) levels were reduced in strains harboring MacP and PBP1A. (B) Representative streaks of the indicated strains on blood agar plates in the absence and presence of 200 μ M ZnCl₂. With the exception of wild-type (WT), the other strains lack pbp1a and pbp2a and harbor the Pzn promoter fused to pbp2a, halo-pbp2a, or halo-pbp2a(A77T). Expression of Halo-PBP2a variants supports growth in strains lacking pbp1a and pbp2a. (C) Representative image of SDS-PAGE gel in which cells harboring Halo-PBP2a variants were labeled with Janelia fluor 549 and lysates resolved by SDS-PAGE and imaged directly using the Typhoon FLA9500. Cells grown in the absence or presence of 200 μ M ZnCl₂ were incubated with the Janelia fluor 549 (50nM for 15 min) prior to lysis. (D) InstantBlue-stained gel of the purified His-PBP2a variants used in the *in vitro* glycosyltransferase assay. The catalytic mutants in the transpeptidase domain (S410A) and the glycosyltransferase domain (E131A) are labeled TP- and GT-, respectively. (E) Representative blot of glycan strand polymerization assay using purified proteins and lipid II. The reaction products and remaining substrate were labeled with

biotinylated D-lysine using *S. aureus* PBP4, resolved by SDS-PAGE, transferred to PVDF, and detected by IRDye 800CW Streptavidin. Free lipid II and glycan strands (PG) are indicated. Some PBP4 and PBP2a become biotinylated in the labeling reaction and are detected by IRDye 800CW Streptavidin. Timepoint: 2 min. A final concentration of 0.5 μ M of PBP2a variants and no cephalixin were used for this specific control.

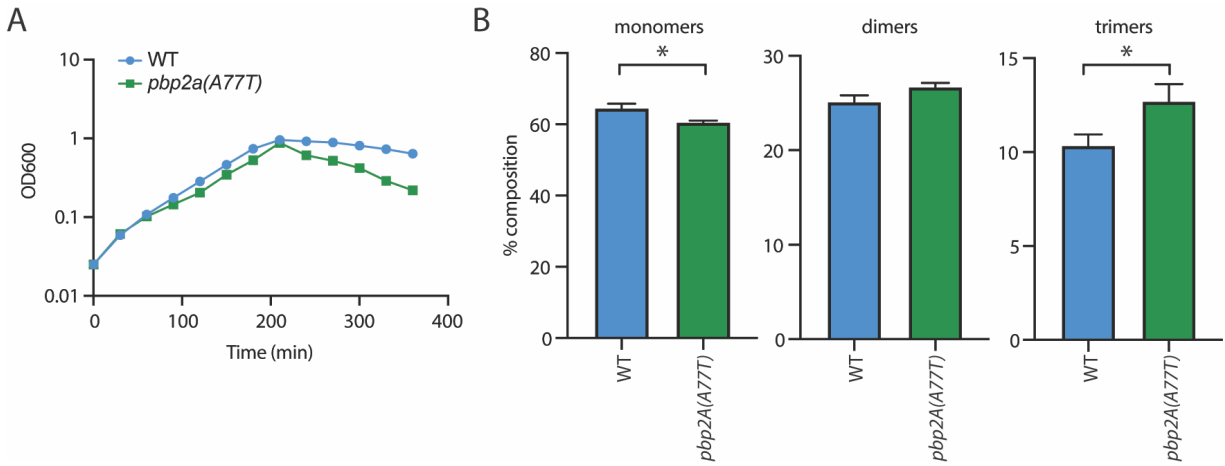


Figure S4 Analyses of strains harboring *pbp2A*(WT) and *pbp2A*(A77T)
(A) Growth curve in THY medium of the indicated strains. **(B)** Bar graph shows the percentage of cell wall crosslinking in the indicated strains. The difference is statistically significant (*) with a *p-value* < 0.05 (*p-value*= 0.018 for the monomers plot; *p-value*= 0.048 for the trimers plot). Monomers, dimers, and trimers refer to uncrosslinked, singly crosslinked, or doubly crosslinked muropeptides detected in the muropeptide analysis of *Sp* peptidoglycan.

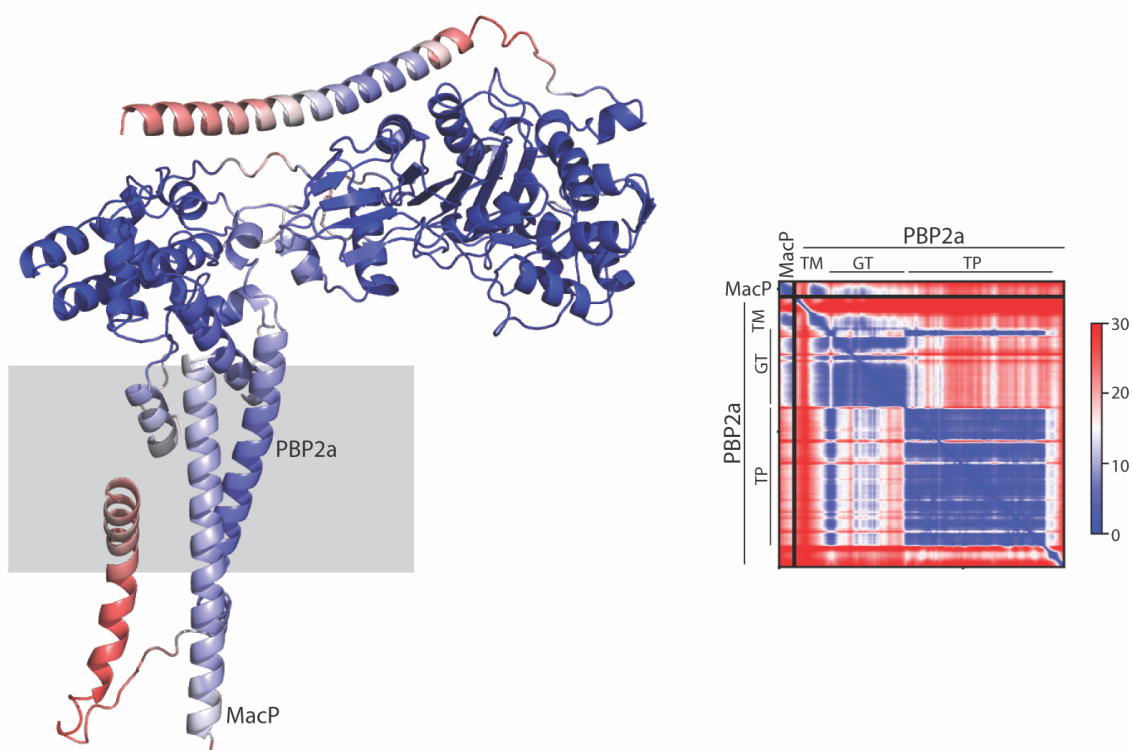


Figure S5: Predicted local difference test and alignment error for the AlphaFold-predicted complex of PBP2a and the transmembrane segment of MacP. Structural model of the PBP2a and the transmembrane segment of MacP with predicted local distance difference tests (pLDDT) per position mapped onto predicted complex (left). Red-white-blue colors indicate low to high confidence. The predicted alignment error in Å of all residues against all residues for the top-ranked model is shown on the right. Given that the extreme N-terminus of PBP2a is unlikely to fold back into the membrane as predicted by AlphaFold, we eliminated this portion of the protein structure from the models shown in other figures for clarity.

Table S1

PBP2a suppressor number	number of mutations	change(s)
3a	4	A167V, G577A, T593T, A699V
29a	6	A105T, Y136Y, N331K, K392K, S404A, E705V
30a	2	A77T, P375S
39a	1	A77T
41a	2	A77T
44a	2	D84N, T571S
4b	1	E325D
9b	4	D84G, N342N, L477F, A719V
14b	2	A77T, P375S
17b	4	L73S, G385A, S535C, P653P
21b	1	A77T
4c	2	A77T, P375S
14c	2	A77T, P375S
16c	-	T571S, poor sequencing
56c	2	K183I, P673L

TableS1: Isolated variants of PBP2a that promote growth upon PBP1a depletion in absence of MacP. Red: A77T mutation with the highest occurrence; blue: substitutions also tested as a single change (D84K and K183I) in validation studies.

Table S2

WT	<i>D39 Δcps</i>	lab collection
6	<i>D39 Δcps, Δpbp2a::Erm</i>	Fenton A. et al. 2018
9	<i>D39 Δcps, Δpbp1a::Kan</i>	Fenton A. et al. 2016
30	<i>D39 Δcps, Δpbp1a::Kan, ΔbgaA::Pzn-pbp1a-Tet, Δpbp2a:Erm</i>	Fenton A. et al. 2018
36	<i>D39 Δcps, Δpbp1a::Kan, ΔbgaA::Pzn-pbp1a-Tet</i>	Fenton A. et al. 2018
94	<i>D39 Δcps, ΔmacP::Spec</i>	Fenton A. et al. 2018
106	<i>D39 Δcps, Δpbp1a::Kan, ΔmacP::Spec, ΔbgaA::pZn-pbp1a-Tet</i>	Fenton A. et al. 2018
1047	<i>D39 Δcps, Δpbp1A::kan, ΔbgaA::Pzn-pbp1A:tet, ΔmacP::Spec, Δpbp2a:erm</i>	This study
1064	<i>D39 Δcps, Δpbp1A::kanR, ΔbgaA::Pzn-pbp1A-tetR, ΔmacP::SpecR, pbp2a(A77T)-CmR</i>	This study
1074	<i>D39 Δcps, Δpbp2a::pbp2a(A77T)-Cm</i>	This study
1075	<i>D39 Δcps, Δpbp2a::Erm, ΔmacP::Spec</i>	This study
1082	<i>D39 Δcps, ΔmacP::Spec, Δpbp2a::pbp2a(A77T)-Cm</i>	This study
1083	<i>D39 Δcps, Δpbp2a::pbp2(A77T) CmR, ΔSPD_0876(macP)::SpecR</i>	This study
1086	<i>D39 Δcps, Δpbp1a::Kan, Δpbp2a::pbp2a(A77T)-Cm</i>	This study
1087	<i>D39 Δcps, Δpbp1a::Kan, ΔmacP::Spec, Δpbp2a::pbp2a(A77T)-Cm</i>	This study
1094	<i>D39 Δcps, 1526:PF6-optRBS-CFP:1527 KanR</i>	This study
1095	<i>D39 Δcps, 1526:PF6-optRBS-CFP:1527 KanR, pbp2a(A77T)-CmR</i>	This study
1115	<i>D39 Δcps, Δpbp1A::Kan, ΔbgaA::Pzn-pbp1A-TetR, ΔmacP::Spec, Δpbp2a::pbp2a(A77I)-CmR</i>	This study
1116	<i>D39 Δcps, Δpbp1A::Kan, ΔbgaA::Pzn-pbp1A-TetR, ΔmacP::Spec, Δpbp2a::pbp2a(A77V)-CmR</i>	This study
1117	<i>D39 Δcps, Δpbp1A::Kan, ΔbgaA::Pzn-pbp1A-TetR, ΔmacP::Spec, Δpbp2a::pbp2a(A77P)-CmR</i>	This study
1118	<i>D39 Δcps, Δpbp1A::Kan, ΔbgaA::Pzn-pbp1A-TetR, ΔmacP::Spec, Δpbp2a::pbp2a(V76T)-CmR</i>	This study
1119	<i>D39 Δcps, Δpbp1A::Kan, ΔbgaA::Pzn-pbp1A-TetR, ΔmacP::Spec, Δpbp2a::pbp2a(R92E)-CmR</i>	This study
1120	<i>D39 Δcps, Δpbp1A::Kan, bgaA::Pzn-pbp1A-TetR, ΔmacP::Spec, Δpbp2a::pbp2a(D84K)-CmR</i>	This study
1121	<i>D39 Δcps, Δpbp1A::Kan, ΔbgaA::Pzn-pbp1A-TetR, ΔmacP::Spec, Δpbp2a::pbp2a(R92D)-CmR</i>	This study
1124	<i>D39 Δcps, Δpbp2a::erm, ΔbgaA::Pzn-HALO-pbp2a-tetR</i>	This study
1146	<i>D39 Δcps, Δpbp2a::erm, ΔbgaA::Pzn-HALO-pbp2a-tetR, Δpbp1a::kanR</i>	This study
1148	<i>D39 Δcps, Δpbp2a::erm, ΔbgaA::Pzn-HALO-pbp2a(A77T)-tetR</i>	This study
1150	<i>D39 Δcps, Δpbp2a::erm, ΔbgaA::Pzn-HALO-pbp2a(A77T)-tetR, Δpbp1a::kanR</i>	This study
1158	<i>D39 Δcps, Δpbp1A::Kan, ΔbgaA::Pzn-pbp1A-TetR, ΔmacP::Spec, Δpbp2a::pbp2a(E325D)-CmR</i>	This study
1160	<i>D39 Δcps, Δpbp1A::Kan, ΔbgaA::Pzn-pbp1A-TetR, ΔmacP::Spec, Δpbp2a::pbp2a(K183I)-CmR</i>	This study

Table S2: *Sp* strains used for this study.

Table S3

plasmid	genotype	Reference
pCCM60	ori pACYC, Δpbp2a: Pnative-pbp2a-cat, bla	This study
pCCM64	ori pACYC, Δpbp2a: Pnative-pbp2a(A77T)-cat, bla	This study
pMFS8	ori pBR/colE1, kanR, lacIq, PT7:His- <i>pbp2a</i>	This study
pCCM97	ori pBR/colE1, kanR, lacIq, PT7:His- <i>pbp2a</i> (A77T)	This study
pCCM98	ori pBR/colE1, kanR, lacIq, PT7:His- <i>pbp2a</i> _TP- (S410A)	This study
pCCM99	ori pBR/colE1, kanR, lacIq, PT7:His- <i>pbp2a</i> _GT- (E131A)	This study
pCCM114	ori pBR/colE1, kanR, lacIq, PT7:His- <i>pbp2a</i> (D84K)	This study
Genscript synthesized PBP2a(A77I)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77I)-cat, bla	This study
Genscript synthesized PBP2a(A77V)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77V)-cat, bla	This study
Genscript synthesized PBP2a(A77P)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77P)-cat, bla	This study
Genscript synthesized PBP2a(V76T)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (V76T)-cat, bla	This study
Genscript synthesized PBP2a(R92E)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (R92E)-cat, bla	This study
Genscript synthesized PBP2a(D84K)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (D84K)-cat, bla	This study
Genscript synthesized PBP2a(R92D)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (R92D)-cat, bla	This study
Genscript synthesized PBP2a(A75T)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A75T)-cat, bla	This study
Genscript synthesized PBP2a(A77C)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77C)-cat, bla	This study
Genscript synthesized PBP2a(A77D)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77D)-cat, bla	This study
Genscript synthesized PBP2a(A77E)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77E)-cat, bla	This study
Genscript synthesized PBP2a(A77F)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77F)-cat, bla	This study
Genscript synthesized PBP2a(A77G)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77G)-cat, bla	This study
Genscript synthesized PBP2a(A77H)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77H)-cat, bla	This study
Genscript synthesized PBP2a(A77K)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77K)-cat, bla	This study
Genscript synthesized PBP2a(A77L)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77L)-cat, bla	This study
Genscript synthesized PBP2a(A77M)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77M)-cat, bla	This study
Genscript synthesized PBP2a(A77N)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77N)-cat, bla	This study
Genscript synthesized PBP2a(A77P)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77P)-cat, bla	This study
Genscript synthesized PBP2a(A77Q)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77Q)-cat, bla	This study

Genscript synthesized PBP2a(A77R)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77R)-cat, bla	This study
Genscript synthesized PBP2a(A77S)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77S)-cat, bla	This study
Genscript synthesized PBP2a(A77W)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77W)-cat, bla	This study
Genscript synthesized PBP2a(A77Y)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (A77Y)-cat, bla	This study
Genscript synthesized PBP2a(K78A)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (K78A)-cat, bla	This study
Genscript synthesized PBP2a(K78D)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (K78D)-cat, bla	This study
Genscript synthesized PBP2a(S79A)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (S79A)-cat, bla	This study
Genscript synthesized PBP2a(S79V)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (S79V)-cat, bla	This study
Genscript synthesized PBP2a(T80A)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (T80A)-cat, bla	This study
Genscript synthesized PBP2a(T80V)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (T80V)-cat, bla	This study
Genscript synthesized PBP2a(K90D)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (K90D)-cat, bla	This study
Genscript synthesized PBP2a(D97K)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (D97K)-cat, bla	This study
Genscript synthesized PBP2a(R98D)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (R98D)-cat, bla	This study
Genscript synthesized PBP2a(E99K)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (E99K)-cat, bla	This study
Genscript synthesized PBP2a(E99R)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (E99R)-cat, bla	This study
Genscript synthesized PBP2a(E100K)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (E100K)-cat, bla	This study
Genscript synthesized PBP2a(E100R)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (E100R)-cat, bla	This study
Genscript synthesized PBP2a(K101D)	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (K101D)-cat, bla	This study
pCCM113	ori pACYC, Δpbp2a: Pnative- <i>pbp2a</i> (K183I)-cat, bla	This study
pGD159	pACYC, SPD_1526- PF6 promoter-SCFP3A-aphA-3, SPD_1527	This study

Table S3: Plasmids Table. Plasmids highlighted in grey were ordered and mutagenized by Genscript.

Table S4

Oligonucleotide	Sequence (5'→3')	use
45	GAGGGAGGAAAGGCAGGA	construction pCCM60
46	CGCCGTATCTGTGCTCTC	construction pCCM60
388	TCATGACAGATGAAGCTTGGCAGACAATTGACGGCTTGACGG	construction pCCM60
389	CCAAGCTTCATCTGTCATGACCATTTCTAC	construction pCCM60
390	GAGAGCACAGATACGGCGTCAAAGCCTAGCTTTCTTGTTATAATGGATAAGATG	construction pCCM60
391	TCCTGCCTTTCCCTCCCTCCGCATGCAAGCATCTTAGCGAAATAGATTGACTATCG	construction pCCM60
392	ACTAGTGCAAAGGTGCACACGAATCTGATG	construction pCCM60
393	CATCAGATTCGTGTGCACCTTTTGCCTAGTCAGGTGGCACTTTTCGGGG	construction pCCM60
394	AGGGATGAGGAGTTGTTCCCTCAAG	sequencing pbp2a locus/plasmid
395	GTTCAAGTGGTGGCTCTACCATTAC	sequencing pbp2a locus/plasmid
396	GCAGAGGATGGAACGTTTGCTC	sequencing pbp2a locus/plasmid
397	TCTGTAGCTGACAAGATGACCAGTATGATG	sequencing pbp2a locus/plasmid
398	GCACTTTGATATCGACCCAAGTACCG	sequencing pbp2a locus/plasmid
399	GATTTTCGTTCTATCATGGAATACACAGCTTTC	sequencing pbp2a locus/plasmid
400	GATGATCTTCTTGAGATCGTTTTGGTCTG	sequencing pbp2a locus/plasmid
401	GATCGTTGTCAGAAGTAAGTTGGCCG	sequencing pbp2a locus/plasmid
417	TCCCTCCGCATGCAAGCATC	random mutagenesis pbp2a
418	CCATCAACTAGAGCCCAATATTCTCTAGAAATC	random mutagenesis pbp2a
441	GAATCTATTTGTTTGCTGTAACCAAGTCGACCAATGTCAATG	mutation pbp2a(A77T)
441	GAATCTATTTGTTTGCTGTAACCAAGTCGACCAATGTCAATG	construction pbp2a(A77T)
469	GTTGAGAGAAAAGCGATAGAATTTTTCCTTGCCTTAG	construction pbp2a(K183I)
522	CCAATCAAAGCGTAGTCCTGGTGCTACAATTAAGCCTTTAGTTGTTTATAC	mutation pbp2a_TP- (S410A)
523	AACTTGCAGAATGCTGTTATTGCGACAGCAGACCGTTCTTTCATAAAAATGACGG	mutation pbp2a_GT- (E131A)
545	CAAGTCGACCAATGTCAATAAATTGCAAAATGCCTTGAAAACTCGG	construction pbp2a(D84K)

Table S4: Oligonucleotides/primers table

Table S5

strain	genotype	Reference
DH5 α λ pir	endA1 hsdR17 glnV44 (= supE44) thi-1 recA1, gyrA96 relA1 ϕ 80dlac Δ (lacZ)M15 Δ (lacZYAargF), U169 zdg-232::Tn10 uidA::pir+	Gibco BRL
Rosetta2(DE3)	F- ompT hsdSB(rB- mB-) gal dcm (DE3), pRARE2 (CamR)	Novagen
MMH594	Enterococcus faecalis EnGen0310	Huycke et al. 1991

Table S5: Other bacterial strains. DH5 α λ pir and Rosetta2(D33) are *E.coli* strains respectively used for cloning and proteins expression. MMH594 is an *Enterococcus faecalis* strain used for purification of lipid II.